



LDH as a risk factor for IHD

Ataf Rashad Ashour

Submitted: 01-02-2022

Revised: 12-02-2022

Accepted: 16-02-2022

I. INTRODUCTION

In the global health setting, trauma is identified as the primary cause of mortality and impairment among the individuals (Johansson et al., 2011), where about 40% of the trauma fatalities occur due to hemorrhage. Gayet-Ageron et al (2018) have established it as the major cause of death which can be potentially prevented. In case, the death is not immediate, then this accounts to increase in the exsanguination, bleeding and prolonged shock which jeopardize the functioning of the organ and results in late mortality (Moffatt, Mitchell and Walke, 2018). Several studies have indicated the development of the scoring systems which practice triage regulation for assessing the various combination of the anatomical injuries along with physiological and laboratory parameters (van Wessem and Leenen, 2018). These scoring systems help in predicting the outcome of the traumatic patients.

Alarhayem et al. (2016) have indicated that episodes of death due to trauma have instilled the interest of various researches who have highlighted setting in which it can occur; encompassing pre-hospital period, on admission to the emergency department (ED), or during hospitalization. Various studies have asserted that researches will continue to explore the traumatic patient's death, in order to identify the source or reason which this mortality based on the fact that these can be potentially treated and prevented (Pfeifer, et al., 2009). It has been asserted by multiple studies that death of the traumatic patients is affected by various factors constituting of age, injury severity, definitive care time, site of injury, care quality, and presence of coagulopathy as well as a haemorrhagic shock. This mortality ratio can be curtailed down by instigating educational programs or avenues for developing the necessary understanding of the individuals for the first aid of the multiple traumas. Besides these cases, there are various predictors which can be controlled at the scene such as upon the patient's arrival in the emergency unit or following his hospitalization.

Generally, when the supply of the oxygen to the vital organ is inadequate, or a state of insufficient perfusion prevails then such a condition is termed as the haemorrhagic shock.

This occurs because of low blood volume and impaired cardiac preload. Reflecting upon the pre-hospital setting, the clinical experience is generally used for the monitoring and shock resuscitating ion of the traumatic patients, along with their assessment and presence of few primary parameters constituting of consciousness level, heart rate and quality, blood pressure and capillary filling time. In case, the evaluation of the fundamental parameters is found to be normal; there still prevails a presence of shock at the organ level or that on the cellular. Jousi et al. (2010) emphasize the critical evaluation of the shock resuscitation at the end, as well as shock resuscitation as whole, must be restricted to certain cases such as when the anaerobic metabolism and tissue acidosis have been successfully reversed. The same study identified the disturbed microvascular perfusion normalization and oxygen supply as the therapeutic factors which thwart the multiple organ failure (MOF) development.

Arterial blood gas (ABG) is recognized as a significant marker for trauma patient's occult poor perfusion and coagulation profile at admittance (Vohra and Paxton, 2013). It is termed as a screening tool for predicting the outcomes which may occur in severely injured patients. Mujuni et al. (2012) have identified six coagulopathy indicators in trauma constituting of the tissue trauma, hypothermia, hemodilution, acidemia, shock, and inflammation. The occurrence of acute traumatic coagulopathy is reported to be found in 28% to 34% of the patients suffering from multiple injuries. Some researchers have demonstrated the patients with coagulopathy are four times more expected to face mortality and eight times more likely to die in the first 24 hours. It is assumed that the findings of the study will help in the rapidly triage of acute trauma patients, along with the identification of high-risk patient who may develop serious complications. On the other hand, it may reduce the use of un-necessary investigations that have no role in the initial assessment of the patients.



II. REVIEW

LDH Structure and its Isoform

LDH is a homo/hetero tetramer made up of four LDHB or LDHA components to form five distinct isoenzymes with variable distributions (Vallender 2019). LDH-1 in cardiac cells, LDH-2 in the reticuloendothelial system, LDH-3 in lung cells, LDH-4 in the pancreas and kidney, and LDH-5 in the rectus and liver muscle are all examples of this isoform. Monocarboxylate transporters allow cells to release and generate lactic acid. The most frequent isoenzyme in muscles is the LDHA homotetramer, often known as the "muscle type" LDH5 or LDH, which prefers to convert pyruvate to NAD⁺ and lactate. The isoenzyme LDHB (homotetramer), also considered the "heart," is the most frequent isoenzyme in the heart and transforms LDH or LDH1, primarily lactate, into NADH and pyruvate. In most tissues, including malignancies, LDH is expressed in a balanced heterotetrameric form (LDH2, LDH3, and LDH4) and a combination of several isoenzymes. Even though LDH is primarily a cytoplasmic enzyme, research has revealed that it can further be identified in mitochondria. In the mitochondrial matrix, mLDH converts L-lactate to pyruvate. According to several studies, a deficiency of LDH isoenzymes hinders mitochondria from oxidizing lactate (Young et al. 2020).

LDH Mechanism Catalytic Activity

The LDH enzymes are activated in a precise order. Because His 195 and Asp 168 remain seem to be critical, forming an LDH / NADH double complex is required before binding to LDH substrates. This is accompanied by a displacement in which a moving pivot loop is created from a collection of surface amino acid sequences in which Arg 109 is hydrogen-bonded to the carbonyl substrate and closed with the substrate-binding pocket to form residues, allowing reagents and components to interact and enzyme catalysis to be facilitated (Andrews and Dyer 2018). LDH activity is regulated by the metabolic shift from aerobic to anaerobic respiration. Transcriptional control, substrate-level control and allosteric control are the three types of regulators that control LDH. The activity of LDH is influenced by the relative concentration and availability of substrates. The enzyme becomes more active as the number of substrates increases with solid muscular action. When the requirement for ATP is higher than the supply of aerobic Pi, AMP, ADP and ATP rise. The physiological capacity of pyruvate dehydrogenase and other pyruvate metabolite enzymes is surpassed when the

glycolytic stream generates pyruvate. Pyruvate and NAD⁺ are transported through LDH in this process, which results in NADH and lactate (Forkasiewicz et al. 2020).

Functions

ATP production by oxidative phosphorylation is at risk when hypoxic or anaerobic cells. This process requires cells to use other metabolites to produce energy. Therefore, LDH is raised in this case to fulfil the energy production requirement. Lactate is constructed by anaerobic glucose conversion; however, it strikes a metabolic dead end. Lactic acid is thus transported to the liver and released into the bloodstream, where lactate is converted to pyruvate by LDH, completing the Cori cycle. Muscle, on the other hand, is depleted in LDH, and oxygen promotes the conversion of pyruvate to lactate. Due to the lack of mitochondria, pyruvate is not further processed in red blood cells but remains in the cytoplasm, where it eventually turns into lactate. In this process, NADH is oxidized to NAD⁺. Glycolysis research requires the occurrence of high intracellular NAD concentrations. Compared to oxidative phosphorylation, which generates ATP per glucose molecule, anaerobic glycolysis generates merely 2 ATP per glucose molecule.

LDH can further catalyze the dehydration of 2-hydroxybutyrate, but its influence on LDH is smaller than that of lactate (Farhana and Lappin 2021; Melkonian and Schury 2019). Lactic acid absorption occurs during periods of hyperglycaemia and physical exertion, which account for 60% of brain metabolism. In cancer cells, the role of LDH, particularly LDHA, is different than in normal cells. Even in oxygen, cancer cells utilize LDH to boost their aerobic metabolism. This phenomenon is known as the Warburg effect (Goodwin et al., 2019). By avoiding oxidative stress on the electron transport chain, abnormal cancer cells gain from switching to anaerobic metabolism. In addition, cancer cells can acquire the metabolic intermediates of the tricarboxylic acid cycle (TCA cycle), which are formed from glucose and pyruvate to generate lipids and nucleic acids to promote prompt cell multiplication.

Association of Pathophysiology with LDH

Since blood LDH isoenzyme levels reflect tissue-specific disease states, LDH testing is significant in clinical practice. LDH can be utilized as an indication of diverse tissue damage due to its isoenzyme form and extensive use. LDH is released into the bloodstream when cells are



injured. This enzyme can remain in the bloodstream for almost seven days, relying on the degree of tissue injury. Because of cytoplasmic loss and increased cell death, elevated serum LDH occurs due to organ depletion (Farhana and Lappin 2021). Acute myocardial infarction (AMI), renal failure, hepatitis, and anaemia are some conditions that can cause tissue damage. LDH is a helpful signal for assessing non-blood body fluids, therapy response, and determining disease severity and prognosis.

In liver injury or AMI, lower LDH levels during treatment imply an effective prognosis and therapeutic reaction. After four days of acute myocardial infarction, the LDH-1 isoenzyme remained high. The concentration of LDH-5 rises when there is liver injury. LDH-5 is much higher than LDH-4 in hepatocellular cancer, such as cirrhosis or hepatitis (Farhana and Lappin 2021). After hydration, elevated levels of LDH are observed in severe bodily fluids, such as pericardial effusions and stomach fluids. As a result, it's commonly used to describe the outflow. In bacterial meningitis, LDH levels in the cerebrospinal fluid increase, while in viral meningitis, they are normal. The percentage of LDH fluid and the upper limit of normal serum LDH (> 0.6) indicate that an inflammatory process is underway, resulting in discharges.

With intracerebral haemorrhage, the concentration of LDH increases significantly. In the central nervous system, lymphomas, leukaemias and metastases grow by more than 40 units / l above normal levels. Augmented levels of several isoenzymes can indicate a range of tissue injury causes, such as when pneumonia is further associated with heart attacks. LDH levels beyond a certain threshold appear to be connected to severe sickness or multiple organ failure. For metastatic melanoma, LDH is the only serum biomarker available. Hypoxia is a common hallmark of malignant tumours because tumour cells demand more oxygen for development than is available. Tumours that produce LDH use mediated energy production to meet the necessity for fast cellular growth resulting, in LDH is a well-known marker of metastasis, specifically in the liver. It is further a good predictor of survival, as people with high LDH have a minor opportunity of survival. The incidence of melanoma metastases can also be predicted using LDH levels. LDH is linked to tyrosine kinase expression in malignancies (Ding et al. 2022; Liu et al. 2020). In most aggressive cancers, the metabolic change from oxidative phosphorylation to greater anaerobic glycolysis (Warburg effect) occurs. This transition is caused

by the overexpression of the LDH-5 isoform, which is present in muscle and liver. Consequently, inhibiting LDH-5 may directly cause tumour development and invasion (Mishra and Banerjee 2019).

LDH is an excellent biomarker to use, mainly since increased LDH levels have been associated with adverse consequences in patients with various viral infections. Recent information in COVID-19 patients reveals that LDH levels differ significantly between severely unwell and non-acutely ill patients (Henry et al. 2020). Erika Poggiali et al. (2020) discovered that in people with CoVID-19, CRP and LDH are associated with respiratory activity (PaO₂ / FiO₂) and might be used to predict respiratory failure. LDH and CRP could be viewed as beneficial tests to identify persons who need closer respiratory observation and more violent supportive therapy at an initial stage to avoid a poor prognosis (Poggiali et al. 2020). In severe COVID-19 patients, LDH was also demonstrated to be a substantial predictor of lung injury (Han et al., 2020). In biological investigations, the effects of cell growth inhibition and cell death have been widely examined. LDH-based cytotoxic assays (Gordon et al. 2018) are the two most frequent techniques for this purpose. When the plasma membrane is damaged, LDH is quickly unconfined into the cell culture fluid, a key component of cell death, necrosis, and other cell destruction. It may be a valuable indicator for cell viability and cell numbers in vitro (Kumar et al. 2018; You et al. 2019). A typical method for assessing cell viability and late cell dispersion is a quantitative investigation of LDH release (Pei et al. 2020). According to Van den Bossche et al. (2020), bacterial disruption of LDH activity caused an underestimate of cytotoxicity in the presence of specific bacterial species.

LDH AS A BIOMARKER OF CANCER

Suppose the underlying heart disease causes the heart muscle to be replaced by capillaries and fibrous tissue. In that case, the LDH distribution of muscle cells can be obscured by the contribution of LDH to these other cells. Surgically precluded hydroxyproline and microscopic analysis have been performed on a small number of tissues. To reduce this possibility, consider looking at the normal contraction of heart tissue and tissue that seemed very normal. Assuming that areas of visible scar tissue are excluded, there is evidence that the proportion of myocardium and collagen remains unchanged regardless of overgrowth, age, ischemia, or sex. It shows the irrespective of pathology or stage of development. Therefore,



based on the limitations of clinical trials, the samples examined showed the LDH composition of the myocardial cells as accurately as possible. Taken from normal human hearts, it shows that the distribution of LDH in the right ventricle and left ventricle is undetectable through the interstitial chamber. The proportion of A-units from these hearts was 12.9%.

This previously analyzed study was not statistically different from the set of control in this series. For anaerobic glycolysis, to remove hydrogen ions from glyceraldehyde-3-phosphate, it must first be transformed to 1-3-diphosphoglycerate. The coenzyme nicotinamide adenine dinucleotide (NAD) is coupled to these hydrogen ions, converting NAD to reduced NADH₂. NADH₂ must be oxidized before the newly generated NAD may accept hydrogen ions from glyceraldehyde-3-phosphate, allowing glycolysis to continue without oxygen. The conversion of pyruvate to lactate occurs in the final step of the glycolysis pathway. LDH catalyzes this reaction by reducing the hydrogen content of NADH₂ in pyruvate, forming lactate, and oxidizing NAD.

So, the critical function of lactic acid makes available a temporary tank to store hydrogen ions until oxygen is available again. The primary purpose of LDH is to control this reservoir, wisely controlling the production and accumulation of lactic acid to ensure a steady and rapid supply of energy but still keep pH within a narrow range of cellular resistance. As different cells' pH tolerance and energy requirements differ due to their activity, their LDH isotopic patterns also differ. For instance, liver and skeletal muscle can easily tolerate comparatively acidic media and need to produce or process large amounts of lactate. This is reflected in high concentrations of LDH-A monomer. According to the recent findings, if a normal heart can convert large amounts of pyruvate into lactate quickly, arrhythmias, heart failure, and sudden death can be expected in healthy individuals. Therefore, the tetramers produced by the myocardium mainly contain the LDH-B subunit. Because excessive lactate buildup effectively inhibits these tetramers, they only convert tiny amounts of pyruvate to lactate.

Ischemic heart failure involves decreased arterial flow sufficient to switch from anaerobic to aerobic metabolism, with associated lactate production. Although the experimental evidence that this occurs under controlled laboratory conditions is overwhelming, the estimated clinical evidence is less convincing. Atrial or exercise fibrillation examines in patients with coronary

heart illness often show reduced lactate production or extraction from the myocardium or delayed venous homeostasis after exercise. Based on reliable experimental data on altered lactose metabolism present in patients who are suffering from coronary heart illness and a possible association of lactate with angina pectoris, long-term exposures to cardiac tissue were confirmed in these studies. Arterial flow to atrial fibrillation but not enough to kill cells anemia does not directly cause a general response, other than renal effects.

Therefore, maintaining the energy supply of ischemic organs becomes the problem at the level of organ. Speaking of the heart, this may primarily occur with lateral growth and the flow of blood to the left ventricle. Once these events turn out to be scarce, there arise changes in the energy transport system of the cell. Since ischemia involves a reduction in the supply of reagents and oxygen, stimulation is given to promote extra use of the glucose which is available. As soon as the chronic anemia lasts, it takes a while to develop enzymes that can facilitate energy transfer under new environmental conditions.

III. CONCLUSION

Since glycolysis cannot proceed without hydrogen ion treatment, and anaerobic alteration of LDH isotopes can be expected. On the other hand, LDH subunit formation is genetically controlled, and despite the fact that the argument which is mentioned above is reasonable in terms of the physiology of environment, it raises a hard biotic query. The query says that, whether a non-proliferating developed cell reprogram its hereditary purpose to adapt to stress under the influence of severe chronic external trauma? This was the query which was asked to recognize the situation existence but this report is not supposed to answer it. However, this is still a mystery regarding the structural transformation of LDH subunits, taking place in the myocardium of the individual suffering from vascular disease, DNA segment instruction of the function of cell though not too condensed that lingering trauma provocations cannot be abolished. An interesting point is that the anaerobic changes first occurred with the increased utilization of glucose and was not based on the cardiac function which was directly impaired due to excessive lactate production. In short, this information suggests that adaptation to general hypoxia is achieved through a systematic and balanced approach that maintains normal levels of oxygen delivery to tissues. This disrupts the cell's normal energy supply. The evidence suggests that fundamental changes occur at the cellular level



when environmental conditions that preclude systemic benefit are excluded. This manifests as a change in the composition of anaerobic enzymes that promote glucose metabolism.

REFERENCES

- [1]. Alarhayem, A.Q., Myers, J.G., Dent, D., Liao, L., Muir, M., Mueller, D., Nicholson, S., Cestero, R., Johnson, M.C., Stewart, R. and O'Keefe, G., 2016. Time is the enemy: mortality in trauma patients with hemorrhage from torso injury occurs long before the "golden hour". *The American Journal of Surgery*, 212(6), pp.1101-1105.
- [2]. Andrews, B.A. and Dyer, R.B., 2018. Small molecule cores demonstrate non-competitive inhibition of lactate dehydrogenase. *MedChemComm*, 9(8), pp.1369-1376.
- [3]. Ding, L., Gosh, A., Lee, D.J., Emri, G., Huss, W.J., Bogner, P.N. and Paragh, G., 2022. Prognostic Biomarkers of Cutaneous Melanoma. *Photodermatology, photoimmunology & photomedicine*.
- [4]. Farhana, A. and Lappin, S.L., 2021. Biochemistry, lactate dehydrogenase. *StatPearls* [Internet].
- [5]. Forkasiewicz, A., Dorociak, M., Stach, K., Szelachowski, P., Tabola, R. and Augoff, K., 2020. The usefulness of lactate dehydrogenase measurements in current oncological practice. *Cellular & Molecular Biology Letters*, 25(1), pp.1-14.
- [6]. Gayet-Ageron, A., Prieto-Merino, D., Ker, K., Shakur, H., Ageron, F.X., Roberts, I., Kayani, A., Geer, A., Ndungu, B., Fawole, B. and Gilliam, C., 2018. Effect of treatment delay on the effectiveness and safety of antifibrinolytics in acute severe haemorrhage: a meta-analysis of individual patient-level data from 40 138 bleeding patients. *The Lancet*, 391(10116), pp.125-132.
- [7]. Goodwin, ML, Pennington, Z., Westbroek, EM, Cottrill, E., Ahmed, AK and Sciubba, DM, 2019. Lactate and cancer: a "lactatic" perspective on spinal tumor metabolism (part 1). *Annals of Translational Medicine*,
- [8]. Gordon, J.L., Brown, M.A. and Reynolds, M.M., 2018. Cell-based methods for determination of efficacy for candidate therapeutics in the clinical management of cancer. *Diseases*, 6(4), p.85.
- [9]. Han, Y., Zhang, H., Mu, S., Wei, W., Jin, C., Tong, C., Song, Z., Zha, Y., Xue, Y. and Gu, G., 2020. Lactate dehydrogenase, an independent risk factor of severe COVID-19 patients: a retrospective and observational study. *Aging (Albany NY)*, 12(12), p.11245.
- [10]. Henry, B.M., Aggarwal, G., Wong, J., Benoit, S., Vikse, J., Plebani, M. and Lippi, G., 2020. Lactate dehydrogenase levels predict coronavirus disease 2019 (COVID-19) severity and mortality: a pooled analysis. *The American journal of emergency medicine*, 38(9), pp.1722-1726.
- [11]. Johansson, P.I., Sørensen, A.M., Perner, A., Welling, K.L., Wanscher, M., Larsen, C.F. and Ostrowski, S.R., 2011. Disseminated intravascular coagulation or acute coagulopathy of trauma shock early after trauma? An observational study. *Critical Care*, 15(6), pp.1-10.
- [12]. Jousi, M., Reitala, J., Lund, V., Katila, A. and Leppäniemi, A., 2010. The role of pre-hospital blood gas analysis in trauma resuscitation. *World Journal of Emergency Surgery*, 5(1), pp.1-7.
- [13]. Kumar, P., Nagarajan, A. and Uchil, P.D., 2018. Analysis of cell viability by the lactate dehydrogenase assay. *Cold Spring Harbor Protocols*, 2018(6), pp.pdb-prot095497.
- [14]. Liu, Y., Wen, L., Chen, H., Chen, Y., Duan, W., Kang, Y., Ma, L., Huang, X. and Lu, J., 2020. Serum lactate dehydrogenase can be used as a factor for re-evaluating first-relapsed multiple myeloma. *Acta Haematologica*, 143(6), pp.559-566.
- [15]. Melkonian, E.A. and Schury, M.P., 2019. Biochemistry, anaerobic glycolysis.
- [16]. Mishra, D. and Banerjee, D., 2019. Lactate dehydrogenases as metabolic links between tumor and stroma in the tumor microenvironment. *Cancers*, 11(6), p.750.
- [17]. Moffatt, S.E., Mitchell, S.J.B. and Walke, J.L., 2018. Deep and profound hypothermia in haemorrhagic shock, friend or foe? A systematic review. *BMJ Military Health*, 164(3), pp.191-196.
- [18]. Mujuni, E., Wangoda, R., Ongom, P. and Galukande, M., 2012. Acute traumatic coagulopathy among major trauma patients in an urban tertiary hospital in sub Saharan Africa. *BMC emergency medicine*, 12(1), pp.1-7.
- [19]. Pei, J., Panina, S.B. and Kirienko, N.V., 2020. An automated differential nuclear staining assay for accurate determination of mitocan cytotoxicity. *JoVE (Journal of Visualized Experiments)*, (159), p.e61295.
- [20]. Pfeifer, R., Tarkin, I.S., Rocos, B. and Pape, H.C., 2009. Patterns of mortality and causes



- of death in polytrauma patients—has anything changed?. *Injury*, 40(9), pp.907-911.
- [21]. Poggiali, E., Zaino, D., Immovilli, P., Rovero, L., Losi, G., Dacrema, A., Nuccetelli, M., Vadacca, G.B., Guidetti, D., Vercelli, A. and Magnacavallo, A., 2020. Lactate dehydrogenase and C-reactive protein as predictors of respiratory failure in COVID-19 patients. *Clinicachimica acta*, 509, pp.135-138.
- [22]. Vallender, E.J., 2019. Genetics of human brain evolution. *Progress in Brain Research*, 250, pp.3-39.
- [23]. Van den Bossche, S., Vandeplassche, E., Ostyn, L., Coenye, T. and Crabbé, A., 2020. Bacterial Interference With Lactate Dehydrogenase Assay Leads to an Underestimation of Cytotoxicity. *Frontiers in Cellular and Infection Microbiology*, p.494.
- [24]. van Wessem, K.J. and Leenen, L.P., 2018. Incidence of acute respiratory distress syndrome and associated mortality in a polytrauma population. *Trauma surgery & acute care open*, 3(1), p.e000232.
- [25]. Vohra, T. and Paxton, J., 2013. Abnormal arterial blood gas and serum lactate levels do not alter disposition in adult blunt trauma patients after early computed tomography. *Western Journal of Emergency Medicine*, 14(3), p.212.
- [26]. You, S.H., Lim, H.D., Cheong, D.E., Kim, E.S. and Kim, G.J., 2019. Rapid and sensitive detection of NADPH via mBFP-mediated enhancement of its fluorescence. *PloS one*, 14(2), p.e0212061.
- [27]. Young, A., Oldford, C. and Mailloux, R.J., 2020. Lactate dehydrogenase supports lactate oxidation in mitochondria isolated from different mouse tissues. *Redox biology*, 28, p.101339.